BRIEF COMMUNICATION

OBSERVATIONS ON THE BREEDING BIOLOGY OF A MICROHYLID FROG (GENUS OREOPHRYNE) FROM NEW GUINEA

The conquest of terrestrial environments is a major theme in vertebrate evolution. Classically the Amphibia have been viewed as an intermediate stage hetween the aquatic and terrestrial groups ^{1,2}. However, the standard view of the amphibian life-cycle involving eggs hatching into aquatic tadpoles which subsequently metamorphose into terrestrial frogs belies the true diversity of reproductive modes in the Amphibia.

The diversity of reproductive modes is greater among amphibians than in other vertebrates³. All orders of the Amphibia include species which deposit eggs in terrestrial environments, and terrestrial development has evolved several times within the order Anura³.

Deposition of eggs out of water is a major step toward terrestriality among amphibians. Therefore, it is not surprising that most variation in reproductive modes among anurans occurs in tropical environments where there are high levels of atmospheric moisture³. A second major step toward terrestriality occurs in those groups which exhibit direct development. The evolution of direct development has been important in the successful invasion of montane environments by amphibians³. Direct development occurs in all Australopapuan microhylid frogs⁴.

Despite their ahundance, little is known about the breeding hiology of New Guinean microhylids. Small numbers of large, heavily yolked eggs are concealed in leaf axils, hollow stems, moss clumps or beneath the ground^{4,5}. They may be attended by an adult frog, usually a male^{4,5}. These frogs generally do not form breeding aggregations (exceptions may be *Asterophrys nurpicula*⁶ and *Sphenophryne mehelyi*⁷) and may call spasmodically. Thus any observations are fortuitous and valuable.

This note reports observations on the reproductive biology of an unidentified species of *Oreophryne* near Tabubil in the Western Province of Papua New Guinea (5°17'S, 141°12'E). This species was assigned to *Oreophryne* on the basis of its partially webbed feet and the presence of small clavicles⁸. Current confusion surrounding the taxonomy of this genus prevents reliable allocation of our specimens to any particular species.

On the evening of 27.xi.1991 a single specimen was found 3.5 km SSW of Tabubil near a small creek, beneath a closed canopy mid-montane rainforest⁹. At the time of collection it was sitting over a clutch of ten eggs on the underside of a leaf 0.8m from the edge of the creek and 1.5m above the ground (Fig. 1). It was not heard calling, but its sex was determined later by dissection. Only two other species of frog were heard calling in the area (*Rana grisea* and an undescribed member of the *Litoria eucnemis* species-group).

The clutch was arranged in two vertical lines of four and six eggs each. The entire clutch was covered with a thin, transparent, memhranous structure which yellowed slightly in alcohol. This layer was quite separate from the jelly-like capsule which surrounded the embryos and, although superficially different, may be homologous to the cord that joins the eggs of congeners for which the eggs are known^{8,10,11}. The capsules were 3.8-4.0mm in diameter. The



Fig. 1. A male *Oreophryne* (SAM R40884) attending a clutch of eggs on the underside of a leaf near Tabubil, Papua New Guinea.

embryos were 3.7-3.8mm long. Each egg contained a very large, ovoid yolk body 3.1-3.2mm in maximum diameter.

A single egg from this clutch was dissected (Fig. 2). The head (1.1×1.1mm) could be distinguished clearly from the laterally compressed body, which was 2.6mm long. Four limb buds were present, although no differentiation of the digits was apparent. Posterior to the hindlimh buds the tail was strongly compressed laterally, but no vascularisation was apparent in the preserved material. Three cephalic lobes and a pair of pigmented eye spots were clearly visible. A single pair of gill buds were present on each side of the head. Dorsally, two dark lines ran the length of the body, indicating that the neural groove had not yet closed. With the exception of the presence of eye pigmentation and the lack of neural groove closure, these microhylid embryos resemble stage five embryos of the South American leptodactylid, *Eleutherodactylus coqui*¹².

Price^{II} illustrated a single, much older embryo of *Oreophryne* from Yapen Island, Irian Jaya. The tail was greatly enlarged and highly vascularised late in development, and presumably plays a role in respiratory gas exchange.

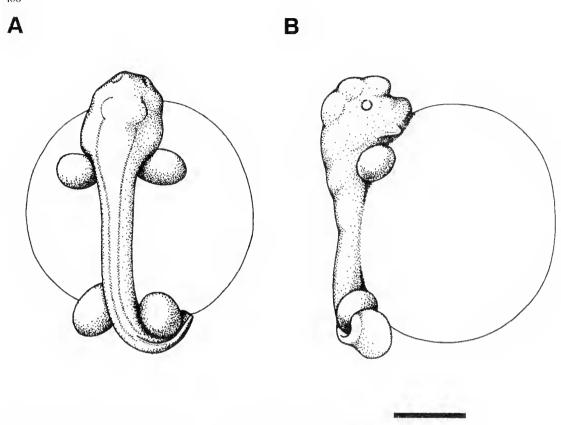


Fig. 2. Dorsal (A) and lateral (B) views of an *Oreophryne* embryo. Scale bar = 1mm.

Measurements of the adult frog were: snout-vent length 22.6mm; head length 7.5mm; head width 7.8mm; eye diameter 3.1mm; eye to naris distance 1.9mm; internarial span 1.8mm; tibia length 10.9mm; foot length 14.2mm. The frog and the clutch it attended have been registered in the South Australian Museum as R40884.

A further two, morphologically identical, male *Oreophryne* were found calling from a cluster of large leaves 3m above the ground, 4m from a sago, swamp amongst mid-montane rainforest 6km SSW of Tabubil at night on 1.xii.1991. Their calls were recorded with a SONY Professional Walkman cassette recorder and an Electret Condenser microphone ECM Z200. Calls were analysed using ULTRASOUND v1.10¹³. The call (Fig. 3) was a biphasic rattle, beginning with a short (0.3-0.4s) series of pulses at 3.2kHz which was followed by a longer (0.7-1.0s) series of pulses at the fundamental and dominant frequency of 2.8kHz, and at 3.2kHz. There were 26 pulses.s⁻¹. Eight calls uttered by two individuals gave a mean call rate of one per 63s. The call length was 1.0-1.4s (mean = 1.1, SD = 0.17).

The breeding behaviour of thirteen of the 83 species of Australopapuan microhylids 14,15,16,17 has been documented 5,10,11,16,18,19,20. Four to 55 eggs are laid 11. This reduction in clutch size associated with large, yolked eggs is a common correlate of terrestrial reproduction in amphibians 21.

Males are known to attend eggs in two *Phrynomantis* spp., two *Cophixalus* spp. and three *Sphenophryne* spp. ^{3,4,(8,19)} Both males and females attend eggs in two *Oreophryne* spp. and one species of *Cophixalus* ^{10,20}. Tyler ⁷ reported aggregations of *Sphenophryne mehelyi* attending their individual clutches of eggs. It is not known how long this association of adult frogs with their clutches may last. The association of frogs with their eggs may function in manipulation of eggs, to provide moisture for terrestrial eggs, in protection against predators or for removal of dead or infected eggs²⁰.

This report is the first record of any direct developing species laying its eggs on the exposed surface of a leaf. A number of indirect developing frogs from other families are known to deposit eggs on vegetation overhanging vater in the neotropics^{22,23}, Africa²⁴, Madasgascar²⁵, the Phillipines²⁶ and New Guinea²⁷. In all of these species the hatching tadpoles fall into the water below. The *Oreophryne* we observed breeding in New Guinea were also found in association with water. This may have been an artefact, however, as we concentrated our searches for frogs around water bodies and did not search the forest surrounding them as thoroughly.

This work was funded in part by grants from the Peter Rankin Trust Fund for Herpetology and the James Cook University. Ok Tedi Mining Ltd, and Phil and Sue Gregory

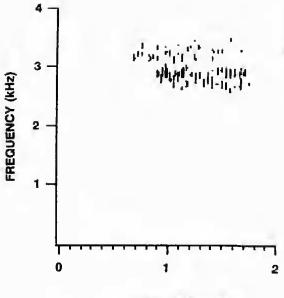


Fig. 3. Sonagram of the call of an Oreophryne from 6 km southeast of Tabubil, Papua New Guinea.

TIME (seconds) ¹⁴Zwelfel, R. G. & Tyler, M. J. (1982) pp. 759-801, In Grissitt, J. L. (Ed.). "Biogeography and ecology of New Guinea." (Junk, The Hague). Carroll, R. L. (1988) "Vertebrate Paleontology and Evolution." (Freeman, New York).

Romer, A. S. (1966) "Vertebrate Paleontology." 3rd Edn (University of Chicago Press, Chicago).

Duellman, W. E. & Trueh, L. (1986) "Biology of the Amphibians." (McGraw-Hill, New York). Burton, T. C. (1990) Trans. R. Soc. S. Aust. 114, 87-93. ¹⁶Blum, P. & Menzles, J. I. (1988) Alytes 7, 125-163. ¹⁷Zweifel, R. G. & Parker, F. (1989) Am. Mus. Novit. 2954, ⁴Zweifel, R. G. (1972) Bull. Amer. Mus. Nat. Hist. 148,

Guinca).

⁵Menzies, J. I. (1976) "Handbook of common New Guinea frogs". (Wau Ecology Institute, Wau). 6S. Donnellan, pers. comm. 1987.

411-546.

⁷Tyler, M. J. (1967) Trans. R. Soc. S. Aust. 91, 187-190. ⁸Parker, H. W. (1934) "A monograph of the frogs of the family Microhylidae." (British Museum (Natural History), London).

⁹Hyndman, D. C. & Menzies, J. I. (1990) J. Biogeog. 17, 241-273.

¹⁰Zweifel, R. G. (1956) Amer. Mus. Novit. 1766, 1-49. Price, D. S. (1992) Sci. New Guinea 18, 3-7.

¹²Townsend, D. S. & Stewart, M. M. (1985) Copeia 1985, 423-436.

¹³Jordan, B. (1988) "Ultrasound V.1,10". (Qucensland University, St Lucia).

assisted with logistic support and transport in New Guinea. Ken Sanderson allowed us to use his software to analyse the call. Nelson Kanem assisted in the field. Margaret Davies and Paula Dempsey made helpful comments on the manuscript. Our work in New Guinea could not have been

done without the help of Guy Kula (Department for the

Environment, PNG), Rosalyn Busava (Institute of Papua New Guinea Studies) and James Menzies (University of Papua New

¹⁸Mehely, L. von (1901) Termes. Fuzetek 24, 169-271. ¹⁹Tyler, M. J. (1963) Trans. R. Soc. S. Aust. 86, 11-29. ²⁰Simon, M. (1983) Behav, Ecol, Sociobiol. 14, 61-67, ²¹Salthe, S. N. & Duellman, W. E. (1973) pp. 229-249, In.

Vial. J. L. (Ed.), "Evolutionary Biology of Anurans: Contemporary Research on Major Problems". (Univ. Missouri Press, Columbia).

²²Bokerman, W. C. A. (1966) Ann. Acad. Brasil. Cien. 38, 335-344.

²³Duellman, W. E. (1970) Monog. Mus. Nat. Hist. Univ. Kansas 1, 1-753.

²⁴Drewes, R. C. (1984) Occ. Pap. California Acad. Sci. 139, 1-70.

²⁵Blommers-Schlosser, R. M. A. (1979) Beaufortia 29, 1-77. ²⁶Alcala, A. C. (1962) Copeia 1962, 679-726.

²⁷Tyler, M. J. (1963) Trans. R. Soc. S. Aust. 86, 105-130.

G. R. JOHNSTON, School of Biological Sciences, Flinders University, G.P.O. Box 2100, Adelaide, S. Aust. 5001, and S. J. RICHARDS, Zoology Department, James Cook University, Townsville, Qld 4811.